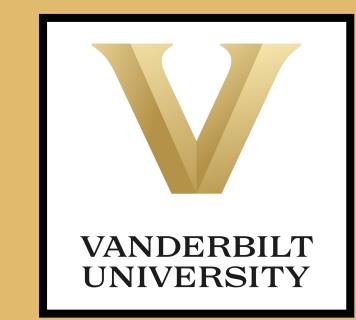


# Shear-thinning, Nanoparticle-based Hydrogels as an Injectable Delivery Platform for Repair of Chronic Diabetic Skin Wounds



Mariah G. Bezold, Andrew R. Hanna, Prarthana Patil, Bryan R. Dollinger, Fang Yu, Mukesh K. Gupta\*, Craig L. Duvall\*

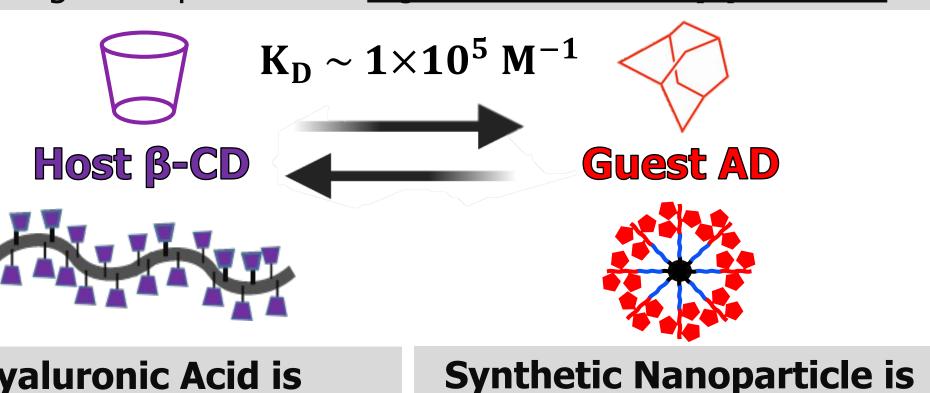
Department of Biomedical Engineering, Vanderbilt University, Nashville, TN

### Introduction

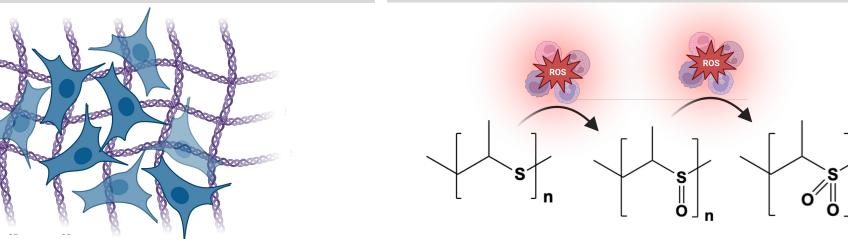
Injectable biomaterials are a promising therapeutic delivery platform for small molecule drugs and therapeutic stem cells, for tissue regeneration and repair applications. 1, 2

### **SHEAR-THINNING HYDROGELS**

formed by non-covalent interactions mediating physical crosslinking show promise as injectable delivery platform.3



### **Hyaluronic Acid is Biologically Active ECM**



### **HA Provides**

Enzymatic degradation Repair and regeneration of damaged tissue<sup>1</sup> Retention of therapeutic cells<sup>2,3</sup>

### **NP Provides**

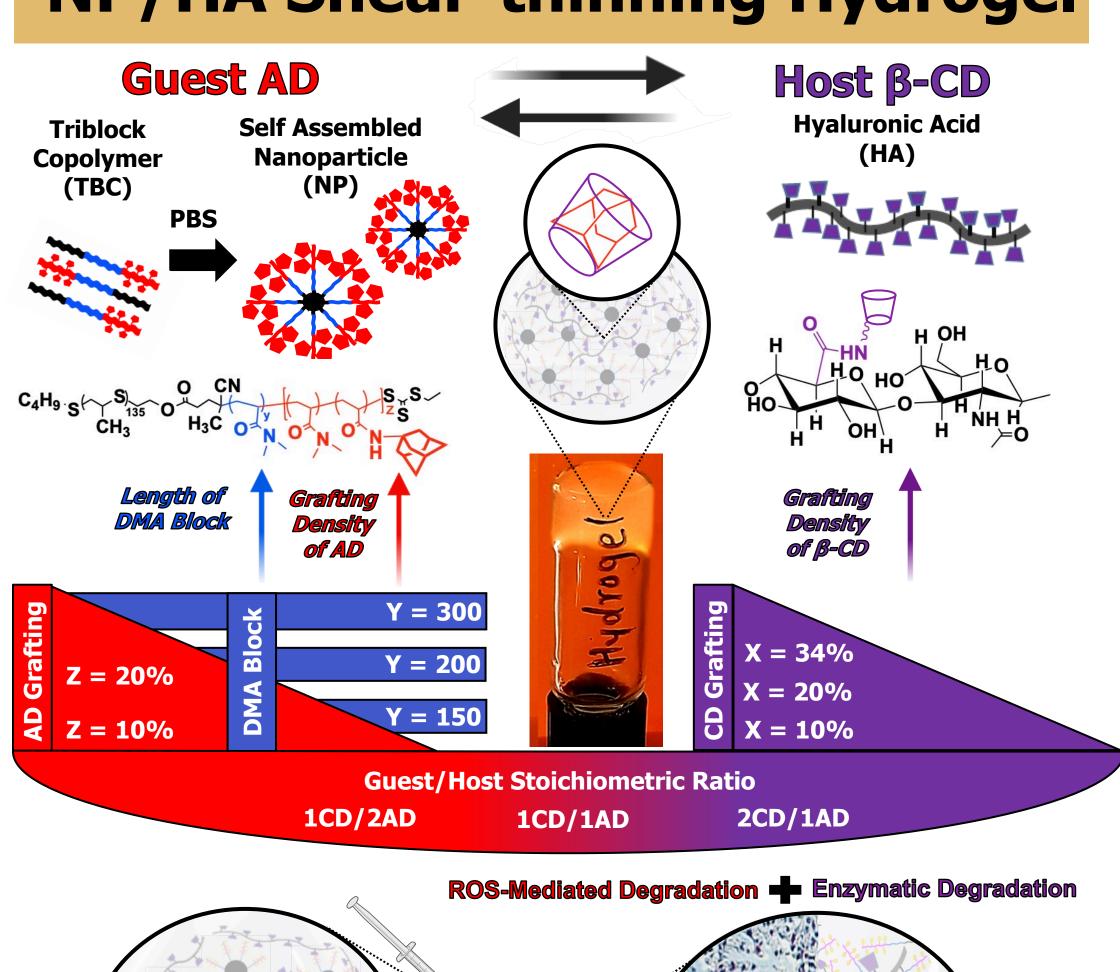
**ROS-Responsive** 

Oxidative degradation<sup>4</sup> Antioxidant behavior ROS-responsive release of small molecule drugs<sup>4</sup>

# **Overall Objective**

Develop NP/HA shear-thinning hydrogel as an injectable, wound-filling platform for sustained delivery of small molecules and retention of therapeutic stem cells to promote repair of chronic diabetic skin wounds

# NP/HA Shear-thinning Hydrogel



### Results

### **Self-Assembly of Stable NPs Grafted with AD**

NP/HA Hydrogel Demonstrates Range of

**Mechanical Strengths and Shear-thinning Behaviors** 

**Figure 2.** (A) Heat map of  $\tan \delta$  for NP/HA hydrogels under low (0.5%) and high

(300%) strain confirms shear-thinning behavior. Effect of AD grafting on G' of

NP/HA hydrogel with increasing DMA blocks (B) 150 units, (C) 200 units, and

(D) 300 units. Effect of CD/AD ratio on G' of NP/HA hydrogel with increasing DMA

NP/HA Hydrogel is Susceptible to

Oxidative and Enzymatic Degradation

PCL D<sub>300</sub>-AD<sub>20%</sub> 1CD/1AD

07 67 10 20 30 50 10 100

PCL D<sub>300</sub>-AD<sub>20%</sub> 1CD/1AD

oxidative (H<sub>2</sub>O<sub>2</sub>), enzymatic (Hyaluronidase), and combined degradation (H<sub>2</sub>O<sub>2</sub> +

Hyaluronidase) shows NP/HA hydrogels are susceptible to both modes of

degradation. Comparison (G-I) G' and (J-L) tan $\delta$  of hydrogels subjected to

PCL D<sub>300</sub>-AD<sub>20%</sub> 1CD/1AD

■ 10 mM H<sub>2</sub>O<sub>2</sub>

1 mM H<sub>2</sub>O<sub>2</sub>-

10 mM H<sub>2</sub>O<sub>2</sub>-

100 mM H<sub>2</sub>O<sub>2</sub>-

blocks and mixed at (E) 1CD/1AD, (F) 2CD/1AD, and (G) 1CD/2AD.

Figure 1. (A) DLS measurements demonstrate A 12 self-assembly of TBCs with varying block length and AD grafting density to form stable NPs. (B) 🤿 Grafting density of 20% for HA-CD results in highest mechanical strength of final NP/HA (C) Cryo-SEM imaging of final NP/HA hydrogel illustrates interconnected porous 3D crosslinked network. Scale bars are 2  $\mu$ m.

D<sub>150</sub>-AD<sub>10%</sub> 1CD/2AD-

D<sub>150</sub>-AD<sub>20%</sub> 1CD/1AD-

D<sub>150</sub>-AD<sub>20%</sub> 2CD/1AD-

D<sub>150</sub>-AD<sub>20%</sub> 1CD/2AD-

D<sub>200</sub>-AD<sub>10%</sub> 1CD/1AD-

D<sub>200</sub>-AD<sub>10%</sub> 1CD/2AD-

D<sub>200</sub>-AD<sub>20%</sub> 1CD/2AD-

D<sub>300</sub>-AD<sub>10%</sub> 1CD/2AD-

D<sub>300</sub>-AD<sub>20%</sub> 1CD/1AD-

D<sub>300</sub>-AD<sub>20%</sub> 2CD/1AD-

**7 U HDEASE** 

■ 10 mM H<sub>2</sub>O<sub>2</sub>

1 mM H<sub>2</sub>O<sub>2</sub>

10 mM H<sub>2</sub>O<sub>2</sub>

100 mM H<sub>2</sub>O<sub>2</sub>

or 6" 18 28 38 58 18 108

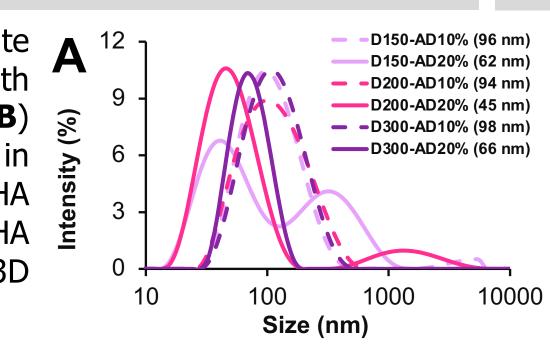
 $D_{300}$ -A $D_{20\%}$  1CD/1AD

Time (hours)

D<sub>300</sub>-AD<sub>20%</sub> 1CD/1AD

degradation of PPS NP/HA hydrogels.

D<sub>300</sub>-AD<sub>20%</sub> 1CD/2AD-



0 6 6 10 20 30 50 10 100

HA/HA 1CD/1AD

Time (hours)

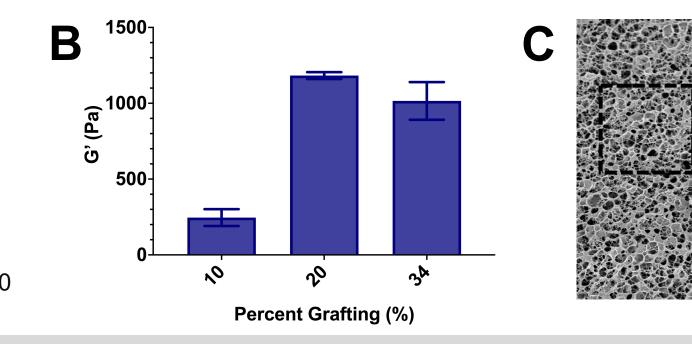
 $1 \text{ mM H}_2\text{O}_2$ 

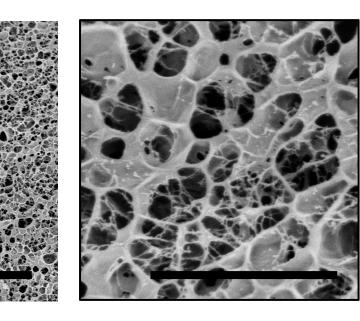
10 mM H<sub>2</sub>O<sub>2</sub>-

100 mM H<sub>2</sub>O<sub>2</sub>-

HA/HA 1CD/1AD

### Synthesis of HA Grafted with β-CD





### NP/HA Hydrogel Provides Injectable Delivery of Cells and Protection from Shear Stress

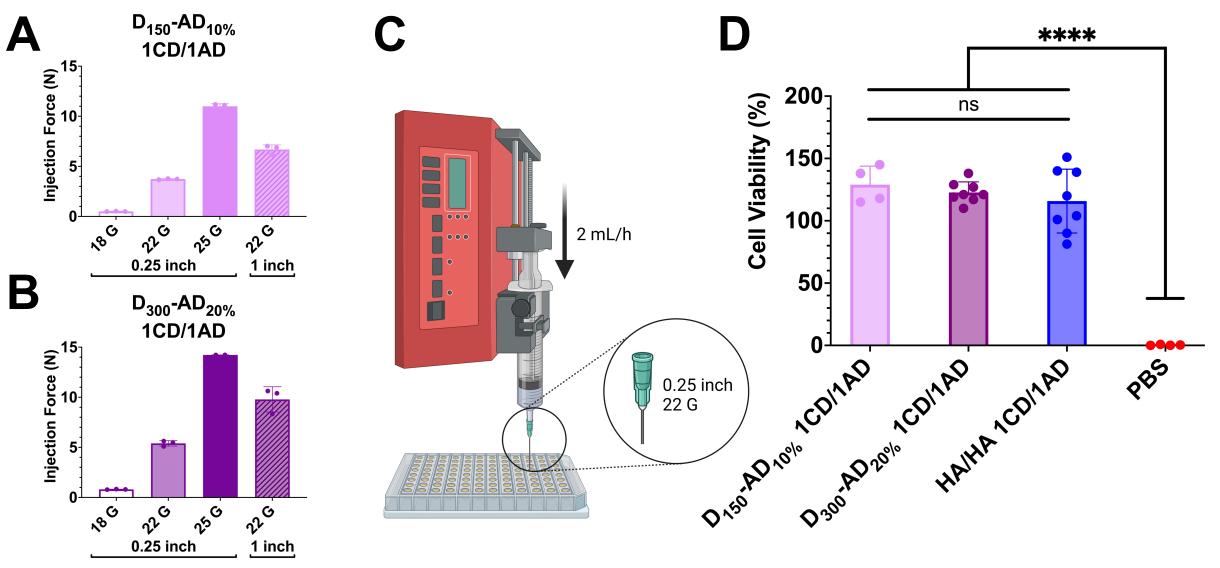


Figure 4. (A-B) Injection force required to pass NP/HA hydrogels through needles of varying gauges and lengths remains below forces acceptable for clinical translation. (C-D) Viability of encapsulated mMSCs following injection at constant flowrate maintained by a syringe pump indicates that NP/HA hydrogels protect cells from shear stress during injection compared to injection of cell suspension in PBS.

### NP/HA Hydrogel is Capable of **Cellular Encapsulation and Protection from ROS**

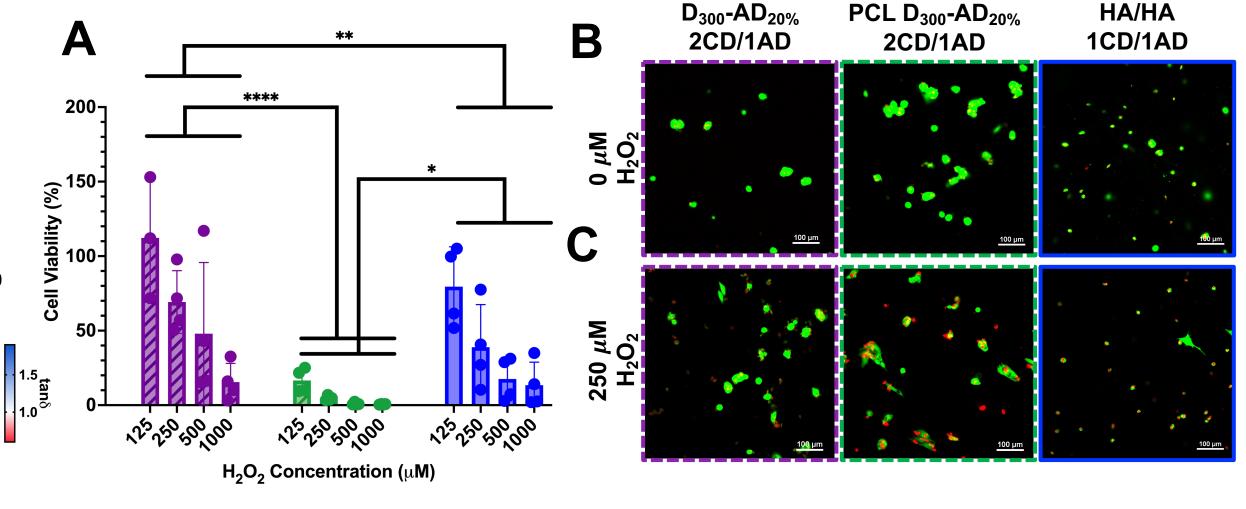
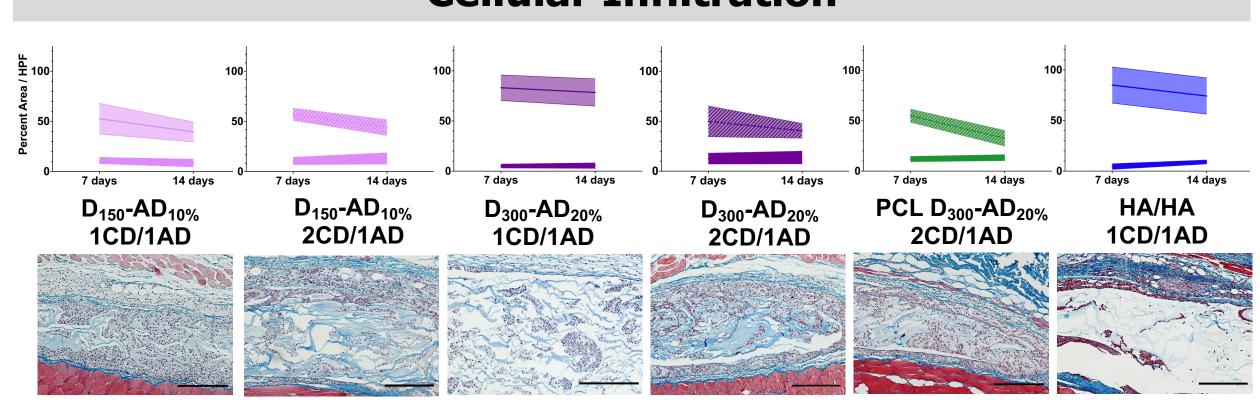


Figure 5. (A-C) Viability of encapsulated mMSCs indicates that NP/HA hydrogels containing PPS provide enhanced viability compared to similar non ROS-responsive shear-thinning hydrogels when subjected to increasing doses of  $H_2O_2$ .

### NP/HA Hydrogel Degradation Profile is Mediated by **Cellular Infiltration**



**Figure 3.** Comparison of (A-C) G' and (D-F) tan $\delta$  of hydrogels subjected to Figure 6. Histological evaluation of excised gel/tissue sections illustrates rate of cellular infiltration (bottom shaded curve) alongside hydrogel degradation (top patterned curve) and hydrogel biocompatibility with surrounding tissue. Scale bars represent 100  $\mu m$ . increasing levels of oxidative degradation (H<sub>2</sub>O<sub>2</sub>) demonstrates ROS-responsive

### Conclusions

### NP/HA SHEAR-THINNING HYDROGELS

Provide **dynamic range** of mechanical properties and shear-thinning behaviors

Allow for **modularity due to synergistic effects** of triblock composition and grafting of guest host complexes

Display **susceptibility to oxidative/enzymatic** degradation

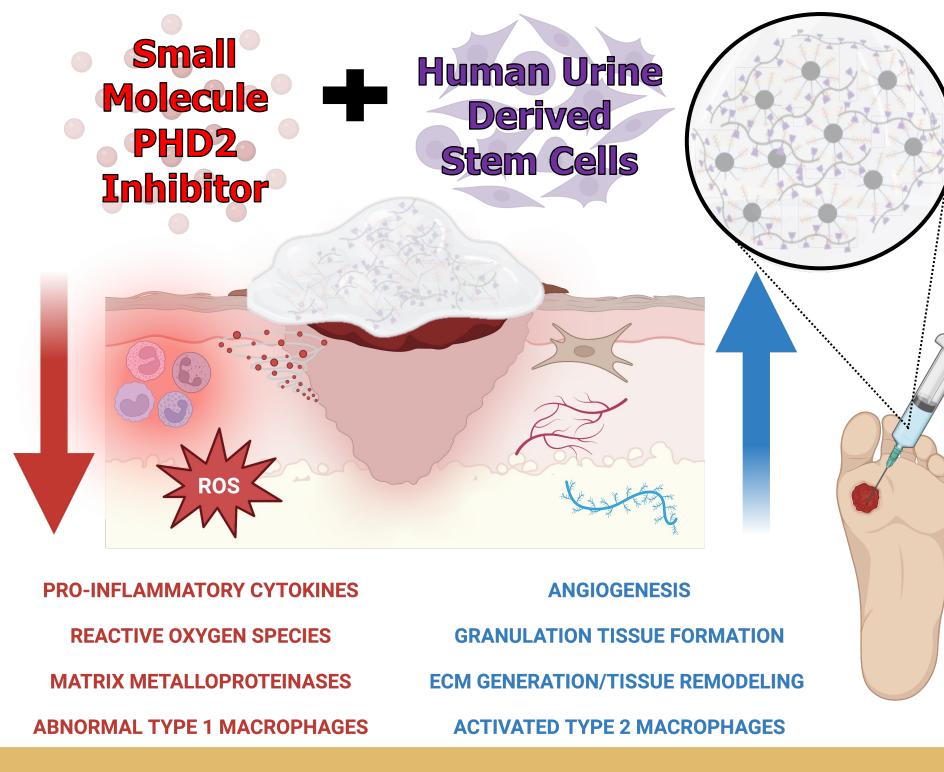
Possess injection forces below forces for **acceptable for** clinical administration

Capable of **protecting encapsulated cells** from cytotoxic ROS and mechanical shear stress

Demonstrate *in vivo* degradation profile mediated by surrounding tissue infiltration

### **Future Work**

Improve synergistic therapeutic outcomes of small molecule PHD2 inhibitor and urine derived stem cells in chronic diabetic skin wounds by providing local delivery and retention in NP/HA hydrogels



# References

- Correa S, Grosskopf AK, Lopez Hernandez H, et al. Translational Applications of Hydrogels. *Chem Rev.* 2021;121(18):11385-11457.
- 2. Muir VG, Burdick JA. Chemically Modified Biopolymers for the Formation Biomedical Hydrogels. 2021;121(18):10908-10949.
- 3. Gaffey AC, Chen MH, Venkataraman CM, et al. Injectable shearthinning hydrogels used to deliver endothelial progenitor cells, enhance cell engraftment, and improve ischemic myocardium. JThorac Cardiovasc Surg. 2015;150(5):1268-1277.
- Gupta MK, Martin JR, Dollinger BR, Hattaway ME, Duvall CL. Thermogelling, ABC Triblock Copolymer Platform for Resorbable Hydrogels with Tunable, Degradation-Mediated Drug Release. Adv Funct Mater. 2017;27(47):1704107.

# Acknowledgements







BioRender

NIH R01 EB028690